Review

Progress in CTLA-4-related immunotherapy for tumors

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Abstract

CTLA-4 (Cytotoxic T Lymphocyte Antigen-4) is an immune checkpoint molecule that can inhibit T cell activation and enhance Regulatery T cell function, thereby reducing the body's immune capacity. In cancer treatment, Ipilimumab, a monoclonal antibody targeting CTLA-4, can effectively inhibit the function of CTLA-4. thereby enhancing the activity and killing ability of T cells. Ipilimumab can also be used in combination with PD-1 and PD-L1 inhibitors for better efficacy. Currently, the FDA has approved Ipilimumab for the treatment of diseases such as melanoma, lung cancer, and colorectal cancer. Although this immunotherapy has shown good efficacy, the side effects and safety issues of the

Correspondence author: Xuelong Jin, Ph.D. degree Professional title: Professor Research direction: Cerebral vascular diseases; Brain science;Nerve regeneration; Microcirculation; Physiology. Department of Physiology and Pathophysiology, Tianjin Medical University, China. treatment need to be highly considered, and further exploration is still needed. Therefore, considering the safety of CTLA-4 therapy, the future direction of immunotherapy mainly involves combining it with other immune checkpoint inhibitors to improve efficacy and reduce side effects.

Keyword e Japan

tumor; mechanism;CTLA-4, PD-1, Ipilimumab, melanoma, non-small cell lung cancer, renal cell carcinoma, colorectal cancer, immune-related adverse events, novel immune checkpoint

1. Background of Immunotherapy

1.1 Overview of CTLA-4 molecule

CTLA-4, also known as cytotoxic T lymphocyte antigen-4, is a member of the immunoglobulin superfamily of adhesion molecules. It is an important co -inhibitory molecule that provides an immune inhibitory signal to T cells. CTLA-4 mainly is expressed on activated T cells after they have produced their effector functions. Upon binding to its ligands CD80 and CD86, CTLA-4 transmits inhibitory signals into the cell, resulting in a reduction in T cell activity and termination of T cell activation. thereby inhibiting the immune response. CTLA-4 is well recognized as а key immune checkpoint[1], and studying its function has led to new progress in the treatment of autoimmune diseases and cancer.

Due to its ability to decrease T cell activity during T cell activation, the CTLA-4 molecule can be used to reduce the immune response in the body. In the treatment of autoimmune diseases, the CTLA-4 molecule is undoubtedly a key pathway and one of the crucial checkpoints for immune checkpoint blockade[2]. In the treatment of autoimmune diseases, a key strategy for the CTLA-4 molecule is to promote its expression, thereby allowing it to bind to the CD80 and CD86 ligands on APCs, reduce the proliferative and differentiating activity of T cells, and thus decrease the immune damage to the body.

1.2 Immune escape mechanism of cancer

The immune escape mechanism of tumors is very complex, closely related to the tumor cells themselves, the microenvironment of tumor growth, and the host immune system.

1.2.1 Tumor cell factors

Firstly, the lack of tumor-specific antigens in tumor cells themselves, and the similarity in structure between the tumor-expressed antigens and normal proteins, weaken the immune recognition and killing ability of the body, and the body cannot produce effective anti-tumor immune responses. At the same time, tumor cells reduce or lose the expression of tumor antigens through antigen modulation, thereby evading recognition and killing by the immune system.

Secondly, the expression level of MHC I molecules on tumor cells themselves is very low, usually cannot reach the number of normal cells, and sometimes even absent, so CTL cannot kill tumor cells.

Thirdly, the co-stimulatory signals of tumor cells are abnormal. Most tumor cells have reduced or missing tumor antigens, and although a small number of tumor cells can express tumor antigens, their surface CD80 and CD86 molecules are lacking, or even high expression of PD-L1 and other coinhibitory molecules, which enables tumor cells to achieve immune escape. This series of abnormal co-stimulatory signals leads to the failure of T cell activation to proceed smoothly, and cannot produce effective immune response to tumor cells.

Fourthly, tumor cells can express or secrete certain immunomolecules that inhibit the body's anti-tumor immune function. Immune molecules such as epidermal growth factor, IL-10, and IL-33 can all play an immunosuppressive role in the body's response to tumors. Tumor cells can even express FasL to induce tumor-specific T cell apoptosis. Fifthly, tumor cells induce the activation and proliferation of Tregs. Tumor cells can actively induce the body to produce Tregs, which can express T cell inhibitory molecules such as LAG-3, GITR, CTLA-4, and PD-1, as well as locally deplete IL-2 and secrete IL-10 and TGF- β [4], thereby allowing tumors to evade the body's immune system attack.

Finally, tumor cells have a certain antiapoptotic effect, which can be achieved by the expression of anti-apoptotic proteins such as Bcl-2 or by the downregulation or mutation of proapoptotic proteins such as BAX.[5] They also have low expression or do not express apoptosis-inducing molecules such as Fas, thereby escaping CTLinduced apoptotic mechanisms.

1.2.2Tumor microenvironmental factors

The main mechanism by which the body forms tumors is that the tumor microenvironment can promote the growth of tumor cells, while also protecting tumor cells from recognition and killing by the body's immune system. The tumor microenvironment is a complex composition, which includes substances that promote tumor cell proliferation and differentiation, as well as components that inhibit the body's immune response, such as Tregs and inhibitory immune factors. The combined action of these substances creates a local microenvironment that is conducive to tumor growth, thus achieving the goal of promoting immune suppression and tolerance to tumorassociated antigens[6].

1.2.3 Host immune function factors

The strength of the host's own immune system cannot be ignored. When the host is suffering from certain diseases such as HIV or long-term use of certain immunosuppressive drugs, the immune system of the body itself will decrease, which will also lead to the recognition and killing of tumor cells by the body's immune system. In addition, tumor cells can also affect the microenvironment. Certain substances expressed by tumor cells that inhibit the immune function of the body can also cause a decrease in the immune function of the body in turn.

Function and mechanism of CTLA 4

The molecular mechanism of CTLA-4 involves a competitive inhibitory action. CTLA-4 has a higher affinity for CD80 and CD86 than CD28, which effectively reduces T cell activation.[7] In cancer treatment, after T cells recognize and proliferate in response to tumor cell antigens presented by APCs, they roam the body in search of cancer cells. Eventually, immune negative regulation is triggered through the T cell receptor (TCR) to shut down the anti-tumor T cell response.[8] Therefore, the main strategy of cancer treatment based on the action of CTLA-4 is to block the binding of CTLA-4 with CD80/CD86, to avoid the generation of immune negative regulation.

The main mechanism of action of CTLA -4 is that it can be promoted by other substances or interact with some other substances. For example, indoleamine 2,3-dioxygenase (IDO), an enzyme produced by myeloid-derived suppressor cells (MDSCs), dendritic cells (DCs), macrophages in the tumor and microenvironment, deplete can tryptophan and inhibit Т cell proliferation tryptophan through

L-kynurenine), catabolites (such as differentiation promoting the and activation of Tregs and CTLA-4 expression. [9] CTLA-4 can also bind to protein phosphatase 2A (PP2A), which controls DNA damage response by regulating ataxia-telangiectasia mutated autophosphorylation/activation (ATM) levels, inducing cell apoptosis. [10] In cells, CTLA-4 is mainly located in the cytoplasm of resting T cells, while the level of CTLA-4 on the cell membrane is higher during T cell activation [1], thus CTLA-4 in the cytoplasm can interact with PP2A to activate ATM, amplify DNA damage response, and induce apoptosis. [11] In addition, during TCR activation, CTLA-4 is upregulated by the Src homology 2 domain-containing protein tyrosine phosphatase-2 (SHP-2) pathway, which contains inhibitory PI3K downstream signals, resulting in acute immune negative regulation of T cells. [12]

In addition to inhibiting T cell activation, CTLA-4 also maintains and controls the function of Tregs, which reduce the body's immune response. CTLA-4 in Treg cells can interact with protein kinase C- η (PKC- η) and recruit GIT2, PIX, and PAK2 to form a complex that depletes APC CD80/86[7]. This reduces the binding of CD80/CD86 on APCs to co-stimulatory molecules on T cells, thereby blocking co-stimulatory signals the interaction between CTLA-4 and PKC- η is crucial for the inhibitory function of Treg cells[13], and a decrease in CTLA-4 expression or function can lead to the development of autoimmune diseases[14].

3.Clinical application of CTLA-4

3.1 Ipilimumab

Monoclonal antibody against CTLA-4 Ipilimumab is a human immunoglobulin G1 (IgG1) monoclonal antibody that acts as an inhibitor of the CTLA-4 immune checkpoint. It binds to CTLA-4 to inhibit CTLA-4 binding to CD80/CD86 and is the first drug to improve overall survival in patients with advanced melanoma [15,16]. Ipilimumab acts indirectly on the immune system by blocking T cell activation inhibition and reducing Treg immunosuppressive activity to enhance anti-tumor immunity. [17,18] Ipilimumab has been used in the treatment of melanoma in clinical trials, and can be used together with nivolumab in the treatment of advanced melanoma, advanced renal cell carcinoma, nonsmall cell lung cancer and other cancers, and good experimental results have been achieved [19-21]. Therefore. Immunocheckpoint inhibitors can be a reasonable and effective treatment for cancer patients. [22]

3.2 Combination therapy

combination of Ipilimumab and anti-PD-1 drugs

Programmed cell death Protein 1 (PD-1) is a transmembrane protein that plays a critical role in suppressing immune response and promoting autoimmune tolerance by initiating antigen-specific T cell apoptosis and inhibiting Treg apoptosis. [23]PD-1 combines with its ligand to play а role, namely programmed cell death ligand 1 (PD-L1). The signal generated by PD-1 will prevent the phosphorylation of key TCR signaling intermediates [24], activate downstream signaling pathways and inhibit T cell secretion of cytokines [25], thus preventing T cell activation and inducing T cell apoptosis. In the tumor microenvironment, the expression of PD -L1 on tumor cells and antigenpresenting cells is abnormally high, so PD-1/PD-L1 is closely related to tumor immune escape [25], and tumor cells can escape immune killing by binding their own overexpression of PD-L1 to PD-1 expressed on antigen-stimulated T cells [26]

Since CTLA-4 and PD-1/PD-L1 are key immune checkpoints that play a key role in immune regulation, current studies have shown that blocking CTLA-4 and PD-1/PD-L1 pathways also plays a great role in cancer treatment. The important idea is to prevent their binding with their own ligands. Thus, T cells can play a role in strengthening the cell killing mechanism of the body's immune system against tumors. Therefore, CTLA -4 and PD-1/PD-L1 are target molecules for the development of therapeutic antibodies [27].

At present, immune checkpoint blocking targeting the inhibitory immune receptors CTLA-4 and PD-1/PD-L1 has become an effective method for cancer treatment [28]. Immunocheckpoint inhibitor anti-ctLA-4 antibody Ipilimumab is mainly used in the treatment of melanoma patients [12]. PD -L1 immune checkpoint inhibitors PD-1 pembrolizumab antibodies and nivolumab commonly are more implicated in colorectal cancer (CRC), melanoma, renal cell carcinoma (RCC), bladder cancer (BC), NSCLC, head and neck cancer, and other cancer types under clinical investigation [29]. In addition, six drugs that target PD-1 and one drug that targets CTLA-4 have been approved for the treatment of different types of cancer, and these immune checkpoint inhibitors have made a huge impact on the clinical treatment of cancer. Survival of cancer patients has been greatly prolonged with monotherapy, but more than 50% of patients do not respond to treatment, and

long-term effects of the drug have been observed in only a small percentage of patients. [30, 31]

Mouse models have shown tumoryassociated high endothelial varices (TA-HEV) to be the primary site of lymphoid entry into the tumor, while analysis of tumor biopsies in 93 patients with metastatic melanoma showed that TA-HEVs demonstrated better response and survival after anti-PD-1 / anti-ctLA-4 combination therapy. [32] In clinical practice, clinical trials of nivozumab and ipilimumab combined with ICI mab have also shown excellent efficacy for patients with melanoma or renal cell carcinoma [33]. The use of monoclonal antibodies prevents the inhibition of T cell activation and induces the release of effector cytokines and cytotoxic particles to activate T cells targeting tumor cells [34]. The combination of CTLA-4 and PD-1/PD-L1 antibodies not only activates, but also effectively the enhances immune response, including against tumor cells, by synergistically blocking Treg-mediated immunosuppression. [35] Immune checkpoint blocking targeting CTLA-4\PD-1/PD-L1 is currently the most promising systematic therapy, which can be used to treat or even cure many types of cancer, not just melanoma patients [36].

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3.3 Practical experience in combination therapy

Treating with Ipilimumab alone has some limitations, [37]Primary and secondary resistance to single agent checkpoint blockade is an emerging problem. [37]Primary and secondary resistance to single agent checkpoint blockade is an emerging problem in daily clinical routine. However, recent studies suggest that a combination of two immune checkpoint inhibitors may primary resistance prevent [38]. nivolumab and pembrolizumab are used commonly clinically in combination with Ipilimumab, and the objective response rate (ORR) for this treatment is significantly higher than that for single antipd-1 [39]. At present, combined drugs have been used to treat melanoma, renal cell carcinoma, nonsmall cell lung cancer and other cancers [40].

4.Treatment of different types of cancer

4.1 Cutaneous melanoma

For cutaneous melanoma, most patients are diagnosed at an early stage, and surgical treatment is usually the preferred treatment [41]. But still, about 10 percent of melanoma patients are diagnosed with advanced melanoma, where the cancer often has metastasized and cannot be treated with surgery. [42] Thus, the use of immune checkpoint inhibitors became a revolutionary event. Early on, ipilimumab, a CTLA-4 inhibitor, was approved by the FDA for the treatment of metastatic melanoma [43], but this drug needs to be modified as patients develop resistance to the drug. Nivolumab and ipilimumab immunocheckpoint inhibitors have been approved for the treatment of advanced melanoma and have shown promising [44]Pembrolizumab results. also improved progression-free survival and overall survival (OS) relative to ipilimumab. [45]. However, at present, these immune checkpoint inhibitors have some adverse effects, need to be highly careful in clinical use.

4.2 Non-small cell lung cancer

Lung cancer can be divided into two categories according the main to histomorphologic characteristics, prognostic and therapeutic significance: small cell carcinoma (SCLC) and nonsmall cell carcinoma (NSCLC), among which non-small cell carcinoma is more common. [46] Most patients with nonsmall cell lung cancer need systemic treatment, and these patients usually have disease recurrence after surgery, and some patients are even inoperable. [47] For resectable NSCLC, there has

been no new progress in treatment except surgical treatment in the past period of time, and the recurrence rate and mortality after surgery will continue to increase with the progression of disease stage. Therefore, the emergence of immunoadjuvant therapy has become a therapeutic means to improve the survival rate of patients with NSCLC. [48] At present, the first-line treatment for advanced NSCLC is nivolumab plus ipilimumab. This method has a longer overall survival than chemotherapy, and the data taken are very satisfactory. [49] And a chemotherapy-free combination of nivolumab and ipilimumab showed a sustained long-term response in patients with advanced NSCLC and tumors with PD-L1 expression greater than or equal to 1% or less at a four-year follow-up. [50] However, it is worth noting that the occurrence of immune-related adverse events still needs to be paid attention to, and more biomarkers should be used to conduct trials to further study the combination of nivolumab and ipilimumab. [51]

4.3 Renal cell carcinoma

Renal cell carcinoma (RCC) is a malignant tumor that is the seventh most common cancer in men and the ninth most common cancer in women. [52]The most common is the clear cell type (70 -- 90%), followed by papillary

(10 -- 15%) and chromophobe RCCs (3 -- 5%)[53]. Therefore, the treatment of clear cell renal cell carcinoma (ccRCC) is the focus. Medical treatment of RCCS has been transitioning from non-specific immunotherapy with cytokines to targeted therapy of vascular endothelial growth factor (VEGF) and now to new immunotherapy drugs. [54] Clear cell (ccRCC), RCC frequently harbors characteristic second-hit loss-of-function mutations in VHL on a background of loss of chromosome 3p, where VHL resides, the mutation causes the mutant cells to secrete more VEGF. And because CCRCCS are highly permeated by immune cells, VEGF tyrosinase inhibitors (TKIs), immune checkpoint blockers (ICBs), and combinations of these drugs are now widely used to treat the disease. [55] Currently, the most effective approved immunotherapy is nivolumab plus ipilimumab in previously untreated and advanced stages of prior treatment

4.4 Colorectal cancer

Colorectal cancer (CRC) is the third most common cancer diagnosed globally and the second leading cause of cancerrelated death [61]. CRC patients can be divided into three groups: microsatellite instabilities - high (MSI-H), microsatellite instabilities - low (MSI-L) and microsatellite stability (MSS). At

present, nivolumab plus ipilimumab can be used in most dMMR/MSI-H mCRC patients. This treatment has a better response than nivolumab alone [62,63]. However, for the majority of pMMR/MSI-L, the application of immunotherapy has not shown a good therapeutic effect, or even efficacy. [64] Currently, immunotherapy is being explored in combination with other therapies, such as ICIs in combination with radiotherapy, ICIs in combination with small molecule TKIs (such as MEK inhibitors and anti-angiogenesis agents), and ICIs in combination with bispectic antibodies. Clinical use of these combination therapies still requires further studies on safety and efficacy. [65]

5. Side effects and safety issues

Although these immune checkpoint inhibitors (ICIs) have achieved good clinical results, they have certain side effects. In addition to certain drug resistance, they may also cause irAEs. The combination of these ICIs may lead to immune system activation and promote the release of inflammatory cytokines by T cells. Such as interferon gamma and tumor necrosis factor, which may lead to excessive extra-tumor inflammation and autoimmunity [66-68]. irAEs caused by ICIs are most common in the skin, gastrointestinal tract, liver

and endocrine system, and often cause pyitis, rash, diarrhea, fatigue, nausea, elevated transaminase or colitis [69,70], and may also cause diabetes, myocarditis and other diseases, but these diseases are relatively uncommon [71]. ICI may also be neurotoxic and cause a range of neurological complications, such as encephalitis, aseptic meningitis, multiple sclerosis, myasthenia gravis and peripheral neuropathy. [72]

According to the General Terminology Standards for Adverse Events (CTCAE), immune-related adverse events are classified into five levels of toxicity: asymptomatic/mild (level 1), moderate (level 2), severe (level 3), lifethreatening (level 4), and fatal (level 5) [73]. However, a recent study showed that different immune checkpoint inhibitors have different safety profiles, and anti-PD-1 drugs generally show a more favorable safety profile compared to anti-CTLA-4 drugs. [74] In untreated melanoma, 27.3% of patients treated with anti-ctLA-4 developed Grade 3 or 4 irAE, while 16.3% of patients treated with anti-PD-1 developed grade 3 or 4 irAE. Combined treatment with antictLA-4 and anti-PD-1 resulted in a significant increase in the incidence and severity of irAEs, with 55% of patients exhibiting high-grade irAEs[75]. In addition, CTLA-4 inhibitor mainly acts on lymphatic organs, while PD-1

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inhibitor is believed to mainly act on tumor microenvironment, resulting in different IRAes. The former mainly causes pituitaritis and colitis, while the latter mainly causes nephritis, pneumonia, thyroiditis, etc. [76,77]

6. Prospect of the future

At present, most patients develop resistance after using anti-PD-1mab and anti-ctla-4mab, resulting in unsatisfactory long-term efficacy. [78] Therefore, finding new immune checkpoints that trigger monoclonal antibodies against them is a new task, Lymphocyte activating gene -3 (LAG-3), T-cell immunoglobulin and the mucin domain containing omega-3 (TIM-3), Tcell immunoglobulin and ITIM domain (TIGIT), T-cell-activated V-domain Ig inhibitor (VISTA), B7 homologous 3 protein (B7-H3), and B and T-cell lymphocyte attenuator (BTLA)) have been shown to be promising new therapeutic targets with potential clinical applications. [79] For some specific diseases, such as advanced liver cancer and colorectal cancer, the combination therapy of ICI with other ICI, TKI, antiVEGF and other drugs should also be actively explored. [80] In addition, attention should be paid to whether CTLA-4 and other molecules are affected by other factors in cancer treatment. For example, studies have shown that intestinal microflora can affect the blocking of CTLA-4 [81], and studies have also shown that short-chain fatty acids (SCFA) can limit and block the anti-tumor effect of CTLA-4 on the body [82]. These factors need to be further studied in the course of treatment.

7.Conclusion

Existing studies have shown that CTLA-4 is an important target for cancer therapy. of CTLA-4 Use immunosuppressant alone is not very satisfactory, and there are some side effects. Therefore, combination therapy become a new direction has of immunotherapy development, such as the study of new immune checkpoints LAG-3, TIGIT, VISTA, B7-H3 and BTLA. In the future, further research is still needed on how to combine drugs to better improve efficacy and reduce side effects, so as to achieve better tumor treatment.

References

[1]Van Coillie S, Wiernicki B, Xu J. Molecular and Cellular Functions of CTLA-4. Adv Exp Med Biol. 2020;1248:7-32. doi: 10.1007/978-981-15-3266-5_2. PMID: 32185705.

[2]Rowshanravan B, Halliday N, Sansom
DM. CTLA-4: a moving target in
immunotherapy. Blood. 2018 Jan
4;131(1):58-67. doi: 10.1182/blood-2017
-06-741033. Epub 2017 Nov 8. PMID:
29118008; PMCID: PMC6317697.

[3]Liu Y, Zheng P. Preserving the CTLA
-4 Checkpoint for Safer and More
Effective Cancer Immunotherapy.
Trends Pharmacol Sci. 2020 Jan;41(1):412. doi: 10.1016/j.tips.2019.11.003.
Epub 2019 Dec 10. PMID: 31836191;
PMCID: PMC7210725.

[4]Seledtsov VI, Goncharov AG, Seledtsova GV. Clinically feasible approaches to potentiating cancer cellbased immunotherapies. Hum Vaccin Immunother. 2015;11(4):851-69. doi: 10.1080/21645515.2015.1009814. PMID: 25933181; PMCID: PMC4514272.

[5]Hassan M, Watari H, AbuAlmaaty A,
Ohba Y, Sakuragi N. Apoptosis and molecular targeting therapy in cancer.
Biomed Res Int. 2014;2014:150845. doi: 10.1155/2014/150845. Epub 2014 Jun 12.
Retraction in: Biomed Res Int. 2020 Aug 28;2020:2451249. PMID: 25013758; PMCID: PMC4075070.

 [6]Arneth B. Tumor Microenvironment.

 Medicina (Kaunas).
 2019 Dec

 30;56(1):15.
 doi:

 10.3390/medicina56010015.
 PMID:

 31906017; PMCID: PMC7023392.

[7]Kim GR, Choi JM. Current Understanding of Cytotoxic Т Lymphocyte Antigen-4 (CTLA-4) Signaling in T-Cell Biology and Disease Mol Cells. 2022 Therapy. Aug 31;45(8):513-521. doi: 10.14348/molcells.2022.2056. Epub 2022 Jul 27. PMID: 35950451; PMCID: PMC9385567.

[8]Ribas A. Wolchok JD. Cancer immunotherapy checkpoint using Science. blockade. 2018 Mar 23;359(6382):1350-1355. doi: 10.1126/science.aar4060. Epub 2018 Mar 22. PMID: 29567705; PMCID: PMC7391259.

[9]Porcellato I, Brachelente C, Cappelli K, Menchetti L, Silvestri S, Sforna M, Mecocci S, Iussich S, Leonardi L, Mechelli L. FoxP3, CTLA-4, and IDO in Canine Melanocytic Tumors. Vet Pathol. 2021 Jan;58(1):42-52. doi: 10.1177/0300985820960131. Epub 2020 Oct 6. PMID: 33021155.

[10]Goodarzi AA, Jonnalagadda JC, Douglas P, Young D, Ye R, Moorhead GB, Lees-Miller SP, Khanna KK. Autophosphorylation of ataxiatelangiectasia mutated is regulated by protein phosphatase 2A. EMBO J. 2004 Nov 10;23(22):4451-61. doi: 10.1038/sj.emboj.7600455. Epub 2004 Oct 28. PMID: 15510216; PMCID: PMC526470.

[11]Yan Q, Zhang B, Ling X, Zhu B,
Mei S, Yang H, Zhang D, Huo J, Zhao Z.
CTLA-4 Facilitates DNA DamageInduced Apoptosis by Interacting With
PP2A. Front Cell Dev Biol. 2022 Feb
24;10:728771. doi:
10.3389/fcell.2022.728771. PMID:
35281086; PMCID: PMC8907142.

[12]Willsmore ZN. Coumbe BGT. Crescioli S, Reci S, Gupta A, Harris RJ, Chenoweth A, Chauhan J, Bax HJ, McCraw A, Cheung A, Osborn G, Hoffmann RM, Nakamura M, Laddach R, Geh JLC, MacKenzie-Ross A, Healy C, Tsoka S, Spicer JF, Josephs DH, Papa S, Lacy KE, Karagiannis SN. Combined anti-PD-1 and anti-CTLA-4 checkpoint blockade: Treatment of melanoma and immune mechanisms of action. Eur J Immunol. 2021 Mar;51(3):544-556. doi: 10.1002/eji.202048747. Epub 2021 Feb 23. PMID: 33450785.

[13]Kong KF, Fu G, Zhang Y, Yokosuka T, Casas J, Canonigo-Balancio AJ, Becart S, Kim G, Yates JR 3rd, Kronenberg M, Saito T, Gascoigne NR, Altman A. Protein kinase C-η controls CTLA-4-mediated regulatory T cell function. Nat Immunol. 2014 May;15(5):465-72. doi: 10.1038/ni.2866. Epub 2014 Apr 6. PMID: 24705298; PMCID: PMC4040250.

[14]Hosseini A, Gharibi T, Marofi F, Babaloo Z, Baradaran B. CTLA-4: From mechanism to autoimmune therapy. Int Immunopharmacol. 2020 Mar;80:106221. doi: 10.1016/j.intimp.2020.106221. Epub 2020 Jan 30. PMID: 32007707.

[15]Carlino MS, Larkin J, Long GV. Immune checkpoint inhibitors in melanoma. Lancet. 2021 Sep 11;398(10304):1002-1014. doi: 10.1016/S0140-6736(21)01206-X. PMID: 34509219.

[16]Karen D. Price, Frank Simutis, Anthony Fletcher, Lila Ramaiah, Rima Srour, John Kozlosky, Jean Sathish, John Engelhardt, Annette Capozzi, James Crona, Courtni Newsome, Jennifer Wheeler, Daniel Szatkowski, Austin Thekkumthala, Bojing Wang, Wendy Freebern, Helen Haggerty, Todd Bunch, Michael Graziano; Abstract LB-B33: Nonclinical safety evaluation of two distinct second generation variants of anti-CTLA4 monoclonal antibody, ipilimumab, in monkeys. Mol Cancer Ther 1 January 2018; 17 (1_Supplement): LB–B33.

[17]Ascierto PA, Marincola FM, Ribas A.
Anti-CTLA4 monoclonal antibodies: the past and the future in clinical application.
J Transl Med. 2011 Nov 13;9:196. doi: 10.1186/1479-5876-9-196. PMID: 22077981; PMCID: PMC3262765.

[18]Yano H, Thakur A, Tomaszewski
EN, Choi M, Deol A, Lum LG.
Ipilimumab augments antitumor activity of bispecific antibody-armed T cells. J
Transl Med. 2014 Jul 9;12:191. doi: 10.1186/1479-5876-12-191. PMID: 25008236; PMCID: PMC4105782.

[19]Wolchok JD, Chiarion-Sileni V, Gonzalez R, Grob JJ, Rutkowski P, Lao CD, Cowey CL, Schadendorf D, Wagstaff J, Dummer R, Ferrucci PF, Smylie M, Butler MO, Hill A, Márquez-Rodas I, Haanen JBAG, Guidoboni M, Maio M, Schöffski P, Carlino MS, Lebbé C, McArthur G, Ascierto PA, Daniels GA, Long GV, Bas T, Ritchings C, Larkin J, Hodi FS. Long-Term Outcomes With Nivolumab Plus Ipilimumab or Nivolumab Alone Versus Ipilimumab in Patients With Advanced Melanoma. J Clin Oncol. 2022 Jan 10;40(2):127-137. doi: 10.1200/JCO.21.02229. Epub 2021

PMC8718224. [20]Albiges L, Tannir NM, Burotto M, McDermott D, Plimack ER, Barthélémy P, Porta C, Powles T, Donskov F, George S, Kollmannsberger CK, Gurney

Nov 24. PMID: 34818112; PMCID:

H, Grimm MO, Tomita Y, Castellano D, Rini BI, Choueiri TK, Saggi SS, McHenry MB, Motzer RJ. Nivolumab plus ipilimumab versus sunitinib for first -line treatment of advanced renal cell carcinoma: extended 4-year follow-up of the phase III CheckMate 214 trial. ESMO Open. 2020 Nov;5(6):e001079. doi: 10.1136/esmoopen-2020-001079. PMID: 33246931; PMCID: PMC7703447.

[21]Owonikoko TK, Park K, Govindan R, Ready N, Reck M, Peters S, Dakhil SR, Navarro A, Rodríguez-Cid J, Schenker M, Lee JS, Gutierrez V, Percent I, Morgensztern D, Barrios CH, Greillier L, Baka S, Patel M, Lin WH, Selvaggi G, Baudelet C, Baden J, Pandya D, Doshi P, Kim HR. Nivolumab and Ipilimumab as Maintenance Therapy in Extensive-Disease Small-Cell Lung Cancer: CheckMate 451. J Clin Oncol. 2021 Apr 20;39(12):1349-1359. doi: 10.1200/JCO.20.02212. Epub 2021 Mar 33683919; 8. PMID: PMCID: PMC8078251.

[22]Naimi A, Mohammed RN, Raji A, Chupradit S, Yumashev AV, Suksatan W, Shalaby MN, Thangavelu L, Kamrava S, Shomali N, Sohrabi AD, Adili A, Noroozi-Aghideh A, Razeghian E. Tumor immunotherapies by immune checkpoint inhibitors (ICIs); the pros and cons. Cell Commun Signal. 2022 Apr 7;20(1):44. doi: 10.1186/s12964-022-00854-y. PMID: 35392976; PMCID: PMC8991803.

[23]Han Y, Liu D, Li L. PD-1/PD-L1
pathway: current researches in cancer.
Am J Cancer Res. 2020 Mar 1;10(3):727
-742. PMID: 32266087; PMCID:
PMC7136921.

[24]Buchbinder EI, Desai A. CTLA-4
and PD-1 Pathways: Similarities,
Differences, and Implications of Their
Inhibition. Am J Clin Oncol. 2016
Feb;39(1):98-106. doi:
10.1097/COC.00000000000239.
PMID: 26558876; PMCID:

PMC4892769.

[25]Jiang Y, Chen M, Nie H, Yuan Y. PD-1 and PD-L1 in cancer immunotherapy: clinical implications and future considerations. Hum Vaccin Immunother. 2019;15(5):1111-1122. doi: 10.1080/21645515.2019.1571892. Epub 2019 Mar 19. PMID: 30888929; PMCID: PMC6605868. [26]Zhang H, Dai Z, Wu W, Wang Z, Zhang N, Zhang L, Zeng WJ, Liu Z, Cheng Q. Regulatory mechanisms of immune checkpoints PD-L1 and CTLA-4 in cancer. J Exp Clin Cancer Res. 2021 Jun 4;40(1):184. doi: 10.1186/s13046-021-01987-7. PMID: 34088360; PMCID: PMC8178863.

[27]Perez-Santos M. Bispecific anti-PD-1/CTLA-4 antibody for advanced solid tumors. Pharm Pat Anal. 2020
Sep;9(5):149-154. doi: 10.4155/ppa-2020-0017. Epub 2020 Sep 22. PMID: 32960139.

[28]Ott PA, Hodi FS, Robert C. CTLA-4 and PD-1/PD-L1 blockade: new immunotherapeutic modalities with durable clinical benefit in melanoma patients. Clin Cancer Res. 2013 Oct 1;19(19):5300-9. doi: 10.1158/1078-0432.CCR-13-0143. PMID: 24089443.

[29]Dermani FK, Samadi P, Rahmani G,
Kohlan AK, Najafi R. PD-1/PD-L1
immune checkpoint: Potential target for
cancer therapy. J Cell Physiol. 2019
Feb;234(2):1313-1325. doi:
10.1002/jcp.27172. Epub 2018 Sep 7.
PMID: 30191996.

[30]Shen X, Zhao B. Efficacy of PD-1 or
PD-L1 inhibitors and PD-L1 expression
status in cancer: meta-analysis. BMJ.
2018 Sep 10;362:k3529. doi:

10.1136/bmj.k3529.PMID: 30201790;-0191-1.PMID: 30546008;PMCID:PMCID: PMC6129950.PMC6292890.

[31]Rotte A. Combination of CTLA-4 and PD-1 blockers for treatment of cancer. J Exp Clin Cancer Res. 2019 Jun 13;38(1):255. doi: 10.1186/s13046-019-1259-z. PMID: 31196207; PMCID: PMC6567914.

[32]Asrir A, Tardiveau C, Coudert J, Laffont R, Blanchard L, Bellard E, Veerman K, Bettini S, Lafouresse F, Vina E, Tarroux D, Roy S, Girault I, Molinaro I, Martins F, Scoazec JY, Ortega N, Robert C, Girard JP. Tumorassociated high endothelial venules mediate lymphocyte entry into tumors and predict response to PD-1 plus CTLA -4 combination immunotherapy. Cancer Cell. 2022 Mar 14;40(3):318-334.e9. doi: 10.1016/j.ccell.2022.01.002. Epub 2022 Feb 3. PMID: 35120598.

[33]Hayashi H, Nakagawa K.
Combination therapy with PD-1 or PD-L1 inhibitors for cancer. Int J Clin Oncol.
2020 May;25(5):818-830. doi: 10.1007/s10147-019-01548-1. Epub
2019 Sep 23. PMID: 31549270.

[34]Darvin P, Toor SM, Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. Exp Mol Med. 2018 Dec 13;50(12):1-11. doi: 10.1038/s12276-018 [35]Tekguc M, Wing JB, Osaki M, Long J, Sakaguchi S. Treg-expressed CTLA-4 depletes CD80/CD86 by trogocytosis, releasing free PD-L1 on antigenpresenting cells. Proc Natl Acad Sci U S A. 2021 Jul 27;118(30):e2023739118. doi: 10.1073/pnas.2023739118. PMID: 34301886; PMCID: PMC8325248.

[36]Blank CU, Enk A. Therapeutic use of anti-CTLA-4 antibodies. Int Immunol. 2015 Jan;27(1):3-10. doi: 10.1093/intimm/dxu076. Epub 2014 Jul
18. PMID: 25038057.

[37]Fuereder, T. Resistance to immune checkpoint inhibitors. Next steps and combinational approaches. memo 12, 123–127 (2019). https://doi.org/10.1007/s12254-019-0493 -6

[38]Billan S, Kaidar-Person O, Gil Z.
Treatment after progression in the era of immunotherapy. Lancet Oncol. 2020
Oct;21(10):e463-e476. doi: 10.1016/S1470-2045(20)30328-4. PMID: 33002442.

[39]Bhave P, Ahmed T, Lo SN, Shoushtari A, Zaremba A, Versluis JM, Mangana J, Weichenthal M, Si L, Lesimple T, Robert C, Trojanello C,

Wicky A, Heywood R, Tran L, Batty K, Dimitriou F, Stansfeld A, Allayous C, Schwarze JK, Mooradian MJ, Klein O, Mehmi I, Roberts-Thomson R, Maurichi A, Yeoh HL, Khattak A, Zimmer L, Blank CU, Ramelyte E, Kähler KC, Roy S, Ascierto PA, Michielin O, Lorigan PC, Johnson DB, Plummer R, Lebbe C, Neyns B, Sullivan R, Hamid O, Santinami M, McArthur GA, Haydon AM, Long GV, Menzies AM, Carlino MS. Efficacy of anti-PD-1 and ipilimumab alone or in combination in acral melanoma. J Immunother Cancer. 2022 Jul;10(7):e004668. doi: 10.1136/jitc-2022-004668. PMID: 35793872; PMCID: PMC9260790.

[40]Antonia SJ, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, Yokoi T, Chiappori A, Lee KH, de Wit M, Cho BC, Bourhaba M, Quantin X, Tokito T, Mekhail T, Planchard D, Kim YC, Karapetis CS, Hiret S, Ostoros G, Kubota K, Gray JE, Paz-Ares L, de Castro Carpeño J, Wadsworth C, Melillo G, Jiang H, Huang Y, Dennis PA, Özgüroğlu M; PACIFIC Investigators. Durvalumab after Chemoradiotherapy in Stage III Non-Small-Cell Lung Cancer. Ν Engl J Med. 2017 Nov 16;377(20):1919-1929. doi: 10.1056/NEJMoa1709937. Epub 2017 Sep 8. PMID: 28885881.

[41]Ross, M.I. and Gershenwald, J.E.
(2011), Evidence-based treatment of early-stage melanoma. J. Surg. Oncol., 104: 341-353.
https://doi.org/10.1002/jso.21962

[42]Leonardi GC, Falzone L, Salemi R, Zanghì A, Spandidos DA, Mccubrey JA, Candido S. Libra M. Cutaneous melanoma: From pathogenesis to therapy J (Review). Int Oncol. 2018 Apr;52(4):1071-1080. doi: 10.3892/ijo.2018.4287. Epub 2018 Feb PMID: 29532857; 27. PMCID: PMC5843392.

[43]Hartman RI, Lin JY. Cutaneous Melanoma-A Review in Detection, Staging, and Management. Hematol Oncol Clin North Am. 2019 Feb;33(1):25-38. doi: 10.1016/j.hoc.2018.09.005. PMID: 30497675.

[44]Weber J, Mandala M, Del Vecchio M, Gogas HJ, Arance AM, Cowey CL, Dalle S, Schenker M, Chiarion-Sileni V, Marquez-Rodas I, Grob JJ, Butler MO, Middleton MR, Maio M, Atkinson V, Queirolo P, Gonzalez R, Kudchadkar RR, Smylie M, Meyer N, Mortier L, Atkins MB, Long GV, Bhatia S, Lebbé C, Rutkowski P, Yokota K, Yamazaki N, Kim TM, de Pril V, Sabater J, Qureshi A, Larkin J, Ascierto PA; CheckMate 238 Collaborators. Adjuvant Nivolumab versus Ipilimumab in Resected Stage III or IV Melanoma. N Engl J Med. 2017 Nov 9;377(19):1824-1835. doi: 10.1056/NEJMoa1709030. Epub 2017 Sep 10. PMID: 28891423.

[45]Robert C, Ribas A, Schachter J, Arance A, Grob JJ, Mortier L, Daud A, Carlino MS, McNeil CM, Lotem M, Larkin JMG, Lorigan P, Neyns B, Blank CU, Petrella TM, Hamid O, Su SC, Krepler C, Ibrahim N, Long GV. Pembrolizumab versus ipilimumab in advanced melanoma (KEYNOTE-006): post-hoc 5-year results from an openlabel. multicentre, randomised, controlled, phase 3 study. Lancet Oncol. 2019 Sep;20(9):1239-1251. doi: 10.1016/\$1470-2045(19)30388-2. Epub 2019 Jul 22. PMID: 31345627.

[46]Rodriguez-Canales J, Parra-Cuentas E, Wistuba II. Diagnosis and Molecular Classification of Lung Cancer. Cancer Treat Res. 2016;170:25-46. doi: 10.1007/978-3-319-40389-2_2. PMID: 27535388.

[47]Imyanitov EN. Iyevleva AG. Levchenko EV. Molecular testing and targeted therapy for non-small cell lung cancer: Current status and perspectives. Hematol. Crit Rev Oncol 2021 Jan;157:103194. doi: 10.1016/j.critrevonc.2020.103194. Epub 2020 Dec 11. PMID: 33316418.

[48]Uprety D, Mandrekar SJ, Wigle D, Roden AC, Adjei AA. Neoadjuvant Immunotherapy for NSCLC: Current Concepts and Future Approaches. J Thorac Oncol. 2020 Aug;15(8):1281-1297. doi: 10.1016/j.jtho.2020.05.020. Epub 2020 Jun 6. PMID: 32522713.

[49]Hellmann MD, Paz-Ares L, Bernabe Caro R, Zurawski B, Kim SW, Carcereny Costa E, Park K, Alexandru A, Lupinacci L, de la Mora Jimenez E, Sakai H, Albert I, Vergnenegre A, Peters S, Syrigos K, Barlesi F, Reck M, Borghaei H, Brahmer JR, O'Byrne KJ, Geese WJ. Bhagavatheeswaran P, Rabindran SK, Kasinathan RS, Nathan FE, Ramalingam SS. Nivolumab plus Ipilimumab in Advanced Non-Small-Cell Lung Cancer. N Engl J Med. 2019 21;381(21):2020-2031. Nov doi: 10.1056/NEJMoa1910231. Epub 2019 Sep 28. PMID: 31562796.

[50]Paz-Ares LG, Ramalingam SS, Ciuleanu TE, Lee JS, Urban L, Caro RB, Park K, Sakai H, Ohe Y, Nishio M, Audigier-Valette C, Burgers JA, Pluzanski A, Sangha R, Gallardo C, Takeda M, Linardou H, Lupinacci L, Lee KH, Caserta C, Provencio M, Carcereny E, Otterson GA, Schenker M, Zurawski B, Alexandru A, Vergnenegre A, Raimbourg J, Feeney K, Kim SW, Borghaei H, O'Byrne KJ, Hellmann MD, Memaj A, Nathan FE, Bushong J, Tran P, Brahmer JR, Reck M. First-Line Nivolumab Plus Ipilimumab in Advanced NSCLC: 4-Year Outcomes From the Randomized, Open-Label, Phase 3 CheckMate 227 Part 1 Trial. J Thorac Oncol. 2022 Feb;17(2):289-308. doi: 10.1016/j.jtho.2021.09.010. Epub 2021 Oct 12. PMID: 34648948.

[51]Reuss JE, Anagnostou V, Cottrell TR, Smith KN, Verde F, Zahurak M, Lanis M, Murray JC, Chan HY, McCarthy C, Wang D, White JR, Yang S, Battafarano R, Broderick S, Bush E, Brock M, Ha J, Jones D, Merghoub T, Taube J, Velculescu VE, Rosner G, Illei P, Pardoll DM, Topalian S, Naidoo J, Levy B, Hellmann M, Brahmer JR, Chaft JE, Forde PM. Neoadjuvant nivolumab plus ipilimumab in resectable non-small cell lung cancer. J Immunother Cancer. 2020 Sep;8(2):e001282. doi: 10.1136/jitc -2020-001282. PMID: 32929052; PMCID: PMC7488786.

[52]Rini BI, Campbell SC, Escudier B.
Renal cell carcinoma. Lancet. 2009 Mar 28;373(9669):1119-32. doi: 10.1016/S0140-6736(09)60229-4. Epub 2009 Mar 5. PMID: 19269025.

[53]Warren AY, Harrison D. WHO/ISUP classification, grading and pathological staging of renal cell carcinoma: standards and controversies. World J Urol. 2018 Dec;36(12):1913-1926. doi: 10.1007/s00345-018-2447-8. Epub 2018 Aug 19. PMID: 30123932; PMCID: PMC6280811.

[54]Barata PC, Rini BI. Treatment of renal cell carcinoma: Current status and future directions. CA Cancer J Clin.
2017 Nov;67(6):507-524. doi: 10.3322/caac.21411. Epub 2017 Sep 29.
PMID: 28961310.

[55]Bi K, He MX, Bakouny Z, Kanodia A, Napolitano S, Wu J, Grimaldi G, Braun DA, Cuoco MS, Mayorga A, DelloStritto L, Bouchard G, Steinharter J, Tewari AK, Vokes NI, Shannon E, Sun M, Park J, Chang SL, McGregor BA, Haq R, Denize T, Signoretti S, Guerriero JL, Vigneau S, Rozenblatt-Rosen O, Rotem A, Regev A, Choueiri TK, Van EM. Tumor Allen and immune reprogramming during immunotherapy in advanced renal cell carcinoma. Cancer Cell. 2021 May 10;39(5):649-661.e5. doi: 10.1016/j.ccell.2021.02.015. Epub 2021 Mar 11. PMID: 33711272; PMCID: PMC8115394.

[56]Motzer RJ, Tannir NM, McDermott DF, Arén Frontera O, Melichar B, Choueiri TK, Plimack ER, Barthélémy P, Porta C, George S, Powles T, Donskov F, Neiman V, Kollmannsberger CK, Salman P, Gurney H, Hawkins R, Ravaud A, Grimm MO, Bracarda S, Barrios CH, Tomita Y, Castellano D, Rini BI, Chen AC, Mekan S, McHenry MB, Wind-Rotolo M, Doan J, Sharma P, Hammers HJ, Escudier B; CheckMate 214 Investigators. Nivolumab plus Ipilimumab Sunitinib versus in Advanced Renal-Cell Carcinoma. Ν Engl J Med. 2018 Apr 5;378(14):1277-1290. doi: 10.1056/NEJMoa1712126. Epub 2018 Mar 21. PMID: 29562145; PMCID: PMC5972549.

[57]Motzer RJ, McDermott DF, Escudier B, Burotto M, Choueiri TK, Hammers HJ, Barthélémy P, Plimack ER, Porta C, George S, Powles T, Donskov F, Gurney H, Kollmannsberger CK, Grimm MO, Barrios C, Tomita Y, Castellano D, Grünwald V, Rini BI, McHenry MB, Lee CW, McCarthy J, Ejzykowicz F, Tannir NM. Conditional survival and long-term efficacy with nivolumab plus ipilimumab versus sunitinib in patients with advanced renal cell carcinoma. Cancer. 2022 Jun 1;128(11):2085-2097. doi: 10.1002/cncr.34180. Epub 2022 Apr 5. PMID: 35383908; PMCID: PMC9543316.

[58]Lai Y, Tang F, Huang Y, He C, Chen C, Zhao J, Wu W, He Z. The tumour microenvironment and metabolism in renal cell carcinoma targeted or immune therapy. J Cell Physiol. 2021 Mar;236(3):1616-1627. doi: 10.1002/jcp.29969. Epub 2020 Aug 11. PMID: 32783202.

[59]Bedke J, Kruck S, Gakis G, Stenzl A, Goebell PJ. Checkpoint modulation--A new way to direct the immune system against renal cell carcinoma. Hum Vaccin Immunother. 2015;11(5):1201-8. doi: 10.1080/21645515.2015.1016657.
PMID: 25912622; PMCID: PMC4514323.

[60]Braun DA, Bakouny Z, Hirsch L, Flippot R, Van Allen EM, Wu CJ, Choueiri TK. Beyond conventional immune-checkpoint inhibition - novel immunotherapies for renal cell carcinoma. Nat Rev Clin Oncol. 2021 Apr;18(4):199-214. doi: 10.1038/s41571 -020-00455-z. Epub 2021 Jan 12. PMID: 33437048; PMCID: PMC8317018.

[61]Phipps O, Brookes MJ, Al-Hassi HO.
Iron deficiency, immunology, and colorectal cancer. Nutr Rev. 2021 Jan 1;79(1):88-97. doi: 10.1093/nutrit/nuaa040. PMID: 32679587.

[62]Johdi NA, Sukor NF. Colorectal
Cancer Immunotherapy: Options and
Strategies. Front Immunol. 2020 Sep
18;11:1624. doi:
10.3389/fimmu.2020.01624. PMID:
33042104; PMCID: PMC7530194.

Jan;92:102134.

[63]Overman MJ, Lonardi S, Wong KYM, Lenz HJ, Gelsomino F, Aglietta M, Morse MA, Van Cutsem E, McDermott R, Hill A, Sawyer MB, Hendlisz A, Neyns B, Svrcek M, Moss RA, Ledeine JM, Cao ZA, Kamble S, Kopetz S, André T. Durable Clinical Benefit With Nivolumab Plus Ipilimumab in DNA Mismatch Repair-Deficient/Microsatellite Instability-High Metastatic Colorectal Cancer. J Clin Oncol. 2018 Mar 10;36(8):773-779. doi: 10.1200/JCO.2017.76.9901. Epub 2018 Jan 20. PMID: 29355075.

[64]Dekker E, Tanis PJ, Vleugels JLA,
Kasi PM, Wallace MB. Colorectal cancer.
Lancet. 2019 Oct 19;394(10207):1467-1480.
doi: 10.1016/S0140-6736(19)32319-0. PMID: 31631858.

[65]Fan A, Wang B, Wang X, Nie Y,
Fan D, Zhao X, Lu Y. Immunotherapy in colorectal cancer: current achievements and future perspective. Int J Biol Sci. 2021 Sep 3;17(14):3837-3849. doi: 10.7150/ijbs.64077. PMID: 34671202; PMCID: PMC8495390.

[66]Hussaini S, Chehade R, Boldt RG, Raphael J, Blanchette P, Maleki Vareki S, Fernandes R. Association between immune-related side effects and efficacy and benefit of immune checkpoint inhibitors - A systematic review and meta-analysis. Cancer Treat Rev. 2021 10.1016/j.ctrv.2020.102134. Epub 2020 Dec 3. PMID: 33302134.

doi:

[67]Olson DJ, Eroglu Z, Brockstein B, Poklepovic AS, Bajaj M, Babu S, Hallmeyer S, Velasco M, Lutzky J, Higgs E, Bao R, Carll TC, Labadie B, Krausz T, Zha Y, Karrison T, Sondak VK, Gajewski TF, Khushalani NI, Luke JJ. Pembrolizumab Plus Ipilimumab Following Anti-PD-1/L1 Failure in Melanoma. J Clin Oncol. 2021 Aug 20;39(24):2647-2655. doi: 10.1200/JCO.21.00079. Epub 2021 May 33945288; 4. PMID: PMCID: PMC8376314.

[68]Zappasodi R, Serganova I, Cohen IJ, Maeda M, Shindo M, Senbabaoglu Y, Watson MJ, Leftin A, Maniyar R, Verma S, Lubin M, Ko M, Mane MM, Zhong H, Liu C, Ghosh A, Abu-Akeel M, Ackerstaff E, Koutcher JA, Ho PC, Delgoffe GM, Blasberg R, Wolchok JD, Merghoub T. CTLA-4 blockade drives loss of Treg stability in glycolysis-low tumours. Nature. 2021 Mar;591(7851):652-658. doi: 10.1038/s41586-021-03326-4. Epub 2021 Feb 15. PMID: 33588426; PMCID: PMC8057670.

[69] Horvat TZ, Adel NG, Dang TO, Momtaz P, Postow MA, Callahan MK, Carvajal RD, Dickson MA, D'Angelo SP, Woo KM, Panageas KS, Wolchok JD, Chapman PB. Immune-Related Adverse Events, Need for Systemic Immunosuppression, and Effects on Survival and Time to Treatment Failure in Patients With Melanoma Treated With Ipilimumab at Memorial Sloan Kettering Cancer Center. J Clin Oncol. 2015 Oct 1;33(28):3193-8. doi: 10.1200/JCO.2015.60.8448. Epub 2015 Aug 17. PMID: 26282644; PMCID: PMC5087335.

[70] Boutros C, Tarhini A, Routier E, Lambotte O, Ladurie FL, Carbonnel F, Izzeddine H, Marabelle A, Champiat S, Berdelou A, Lanoy E, Texier M, Libenciuc C, Eggermont AM, Soria JC, Mateus C, Robert C. Safety profiles of anti-CTLA-4 and anti-PD-1 antibodies alone and in combination. Nat Rev Clin Oncol. 2016 Aug;13(8):473-86. doi: 10.1038/nrclinonc.2016.58. Epub 2016 May 4. PMID: 27141885.

[71]Darnell EP, Mooradian MJ, Baruch EN, Yilmaz M, Reynolds KL. Immune-Related Adverse Events (irAEs): Diagnosis, Management, and Clinical Pearls. Curr Oncol Rep. 2020 Mar 21;22(4):39. doi: 10.1007/s11912-020-0897-9. PMID: 32200442.

[72]Haugh AM, Probasco JC, Johnson DB. Neurologic complications of immune checkpoint inhibitors. Expert

Opin Drug Saf. 2020 Apr;19(4):479-488. doi: 10.1080/14740338.2020.1738382. Epub 2020 Mar 11. PMID: 32126176; PMCID: PMC7192781.

[73]Common Terminology Criteria for Adverse Events (CTCAE). National Cancer Institute. 2017.

[74]Bagchi S, Yuan R, Engleman EG. Immune Checkpoint Inhibitors for the Treatment of Cancer: Clinical Impact and Mechanisms of Response and Resistance. Annu Rev Pathol. 2021 Jan 24;16:223-249. doi: 10.1146/annurevpathol-042020-042741. Epub 2020 Nov 16. PMID: 33197221.

[75]Larkin Chiarion-Sileni Gonzalez R, Grob JJ, Rutkowski P, Lao Cowey CL, Schadendorf D. CD. Wagstaff J, Dummer R, Ferrucci PF, Smylie M, Hogg D, Hill A, Márquez-Rodas I, Haanen J, Guidoboni M, Maio M, Schöffski P, Carlino MS, Lebbé C, McArthur G, Ascierto PA, Daniels GA, Long GV, Bastholt L, Rizzo JI, Balogh A, Moshyk A, Hodi FS, Wolchok JD. Five-Year Survival with Combined Nivolumab and Ipilimumab in Advanced Melanoma. N Engl J Med. 2019 Oct 17;381(16):1535-1546. doi: 10.1056/NEJMoa1910836. Epub 2019 Sep 28. PMID: 31562797.

[76]Judd J, Zibelman M, Handorf E, O'Neill J, Ramamurthy C, Bentota S, Doyle J, Uzzo RG, Bauman J, Borghaei H, Plimack ER, Mehra R, Geynisman DM. Immune-Related Adverse Events as a Biomarker in Non-Melanoma Patients Treated with Programmed Cell Death 1 Inhibitors. Oncologist. 2017 Oct;22(10):1232-1237. doi: 10.1634/theoncologist.2017-0133. Epub 2017 Jun 26. PMID: 28652280; PMCID:

PMC5634771.

[77]Postow MA, Sidlow R, Hellmann MD. Immune-Related Adverse Events Associated with Immune Checkpoint Blockade. N Engl J Med. 2018 Jan 11;378(2):158-168. 10.1056/NEJMra1703481. 29320654.

[78]Weinmann A, Galle PR. Role of immunotherapy in the management of hepatocellular carcinoma: current standards and future directions. Curr Oncol. 2020 Nov;27(Suppl 3):S152-S164. doi: 10.3747/co.27.7315. Epub 2020 Nov 1. PMID: 33343209; PMCID: PMC7739523.

[79]Qin S, Xu L, Yi M, Yu S, Wu K, Luo S. Novel immune checkpoint targets: moving beyond PD-1 and CTLA-4. Mol Cancer. 2019 Nov 6;18(1):155. doi: 10.1186/s12943-019-1091-2. PMID: 31690319; PMCID: PMC6833286. [80]Chen Y, Hu H, Yuan X, Fan X, Zhang C. Advances in Immune Checkpoint Inhibitors for Advanced Hepatocellular Carcinoma. Front Immunol. 2022 Jun 10;13:896752. doi: 10.3389/fimmu.2022.896752. PMID: 35757756; PMCID: PMC9226303.

[81]Vétizou M, Pitt JM, Daillère R, Lepage P, Waldschmitt N, Flament C, Rusakiewicz S, Routy B, Roberti MP, Duong CP, Poirier-Colame V, Roux A, Becharef S, Formenti S, Golden E, Cording S, Eberl G, Schlitzer A, Ginhoux F, Mani S, Yamazaki T, Jacquelot N, Enot DP, Bérard M, Nigou J, Opolon P, Eggermont A, Woerther PL, Chachaty E, Chaput N, Robert C, Mateus C, Kroemer G, Raoult D, Boneca IG, Carbonnel F, Chamaillard M, Zitvogel L. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. Science. 2015 Nov 27;350(6264):1079-84. doi: 10.1126/science.aad1329. Epub 2015 Nov 5. PMID: 26541610; PMCID: PMC4721659.

[82]Coutzac C, Jouniaux JM, Paci A, Schmidt J, Mallardo D, Seck A, Asvatourian V, Cassard L, Saulnier P, Lacroix L, Woerther PL, Vozy A, Naigeon M, Nebot-Bral L, Desbois M, Simeone E, Mateus C, Boselli L, Grivel J, Soularue E, Lepage P, Carbonnel F, Ascierto PA, Robert C, Chaput N. Nature medicine Japan, June 30, 2023, Vol.6, issue1

Systemic short chain fatty acids limit antitumor effect of CTLA-4 blockade in hosts with cancer. Nat Commun. 2020 May 1;11(1):2168. doi: 10.1038/s41467-020-16079-x. PMID: 32358520; PMCID: PMC7195489.

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